



Soybean Rust Spore Identification

An Immunofluorescence
Assay



Development Team

- Ohio State University - *Assay development*
 - Anne Dorrance, Sally Miller, Mike Boehm, Fulya Baysal-Tustas, Melanie Lewis Ivey
- USDA ARS - *Antigen prep, spores*
 - Doug Luster
 - Reid Frederick
- US Navy Medical Research Center - *Antibody development*
 - Jill Czarnecki

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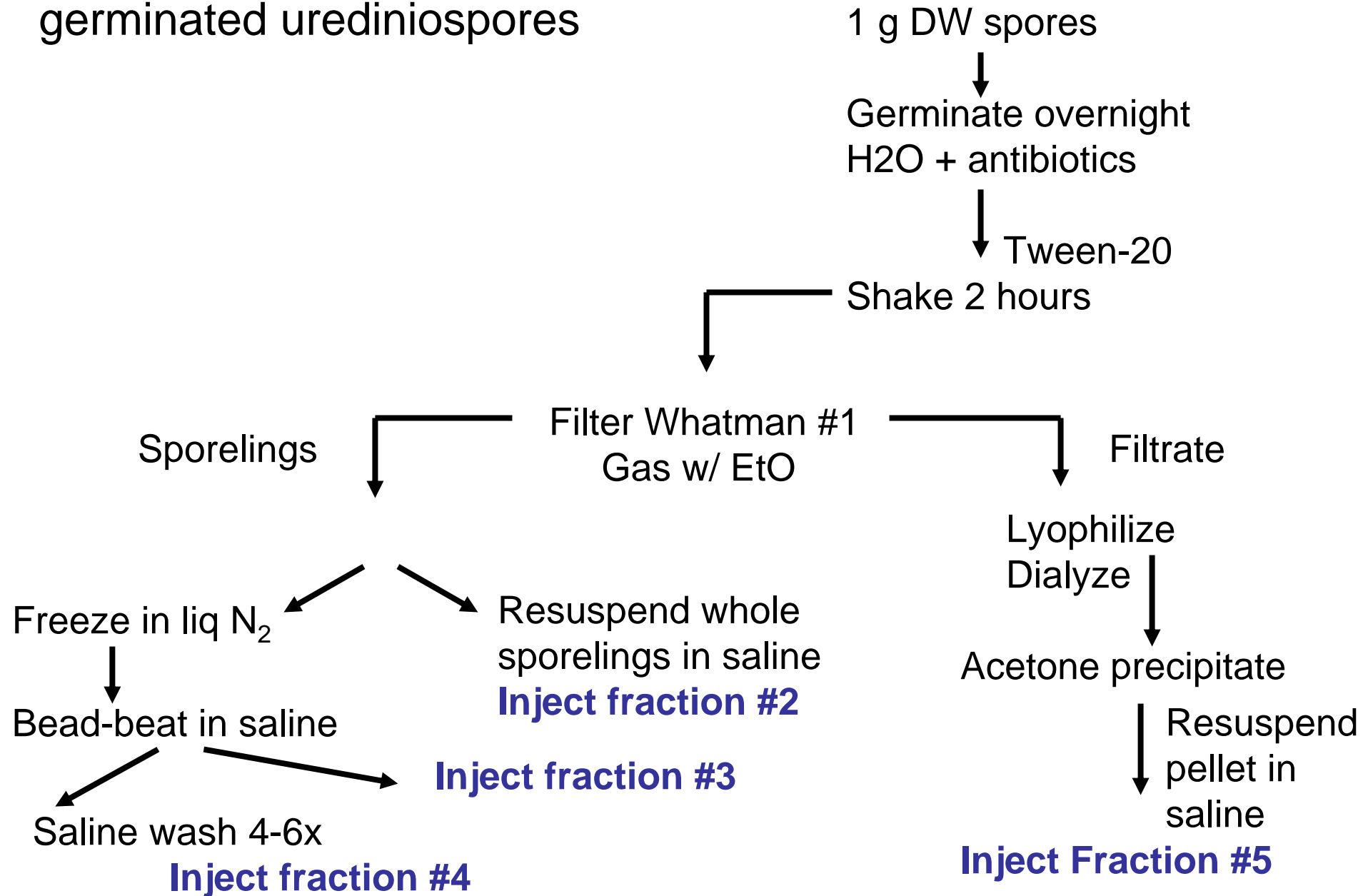
Identification of *Phakopsora* Spores

- Spore monitoring is part of an early warning system
- Spores can be trapped in various air samplers
- *Phakopsora* spores must be identified amidst other spores, etc. in traps
- Immunofluorescence is rapid, simple and definitive

Polyclonal Antibody Development

ID	<i>P. pachyrhizi</i> Antigen Preparation
1A	Ungerminated urediniospores, 20 mg DW/ml
1B	Ungerminated urediniospores, 10 mg DW/ml
2	Intact germ. urediniospores, 1.0 mg protein/ml
3	Pulverized germ. urediniospores, 1.0 mg protein/ml
4	Intact germ. spore walls, 1.25 mg protein/ml
5	Extracellular spore proteins. 0.2 mg/ml

Antigen prep from *P. pachyrrhizi* germinated urediniospores



Specificity of *P. pachyrhizi* Polyclonal Abs

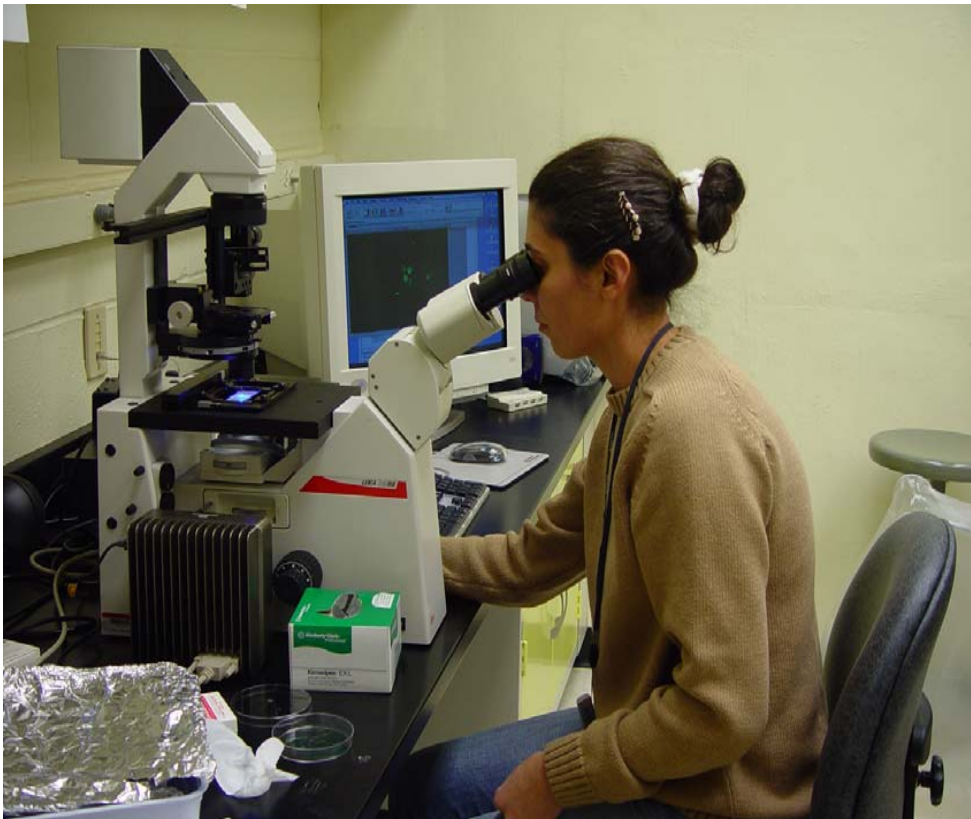
Extract	Reactivity in ELISA				
	1 A	1B	2	3	4
P. pachyrhizi spore extract	+	+	+	+	+
P. meibomia e spore extract	+	+	-	-	-
<i>P. syringae</i> pv. <i>syringae</i> & <i>phaseolicola</i> , <i>X. campestris</i> pv. <i>glycines</i> , <i>C. michiganensis</i> subsp. <i>insidiosus</i> , <i>B. cinerea</i> , <i>C. acutatum</i> , <i>C. gleosporiodes</i>	-	-	-	-	-
<i>Ustilago tritici</i>	-	-	-	-	-
<i>Rhizoctonia solani</i>	-	-	-	+	+
<i>Puccinia graminis</i> f. sp. <i>tritici</i>	-	-	-	-	+
Diseased soybean tissue: bacterial pustule, frog-eye leaf spot, bacterial spot, Septoria leaf spot and powdery mildew	-	-	-	-	-
Symptomless soybean tissue: 183 cultivars	-	-	-	-	-



SBR Immunofluorescence Assay

- Demonstrate specific antibody binding to spores
 - Worked with antibodies 1A and 2
- Adapt to slide capture
- Optimize indirect immunofluorescence assay
- Test using field samples

Epifluorescence Microscopy



Dr. Fulya Baysal-Tustas

- Leica DM IRB microscope
- Optronics Magnafire camera/digital imaging system
- Filter
I3 Blue, Excitation= 450-490, Emission=515
- Exposure: 1.037 sec



Immunoassay Parameters

Parameter	Variable	Optimal result
1° Antibody	Incubation time (1, 2, 3 hr)	2 hr incubation
1° Antibody	Incubation temp: 22°C/37 ° C	22°C
Blocker	2% BSA + 5% goat serum	No blocking step
Washing step	PBS-Tween or water	PBS-Tween
2° Antibody	Incubation time (1, 2, 3 hr)	1 hr incubation
2° Antibody	Incubation temp: 22°C/37 ° C	22°C



Immunofluorescence Assay

Apply 1° Antibody 1A or 2 (1:500) in PBS-Tween



Incubate 2 hr, 22C



Wash 6X, PBS-Tween



Apply 2° Antibody (Alexa Fluor 488, 1:500) in PBS-Tween



Incubate 1 hr, 22 C in dark



Wash 6X, PBS-Tween

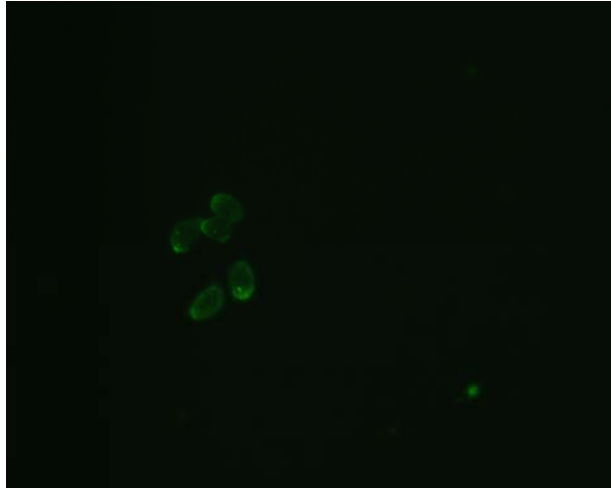


Secondary Antibody

- Alexa Fluor 488 goat anti-rabbit IgG (H+L), 2 mg/ml
 - Excited at 488 nm
 - Maximal emission at 519 nm
 - Brighter and more photostable than FITC

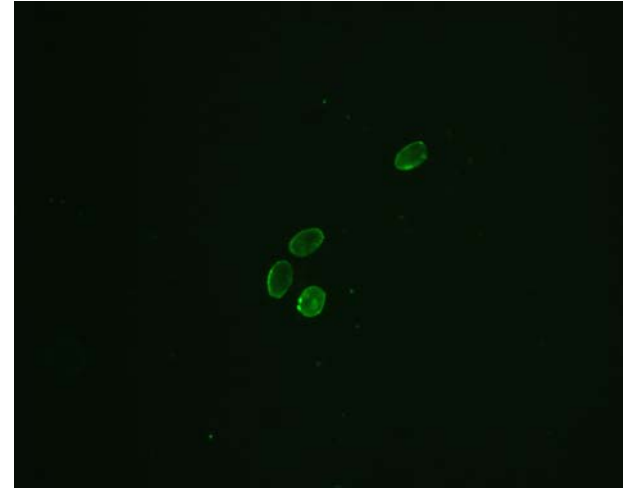
Phakopsora pachyrhizi Immunofluorescence Detection

AB 1A



1:500
1° AB;
1:500
2° AB

AB 2



No 1°
AB;
1:500
2° AB

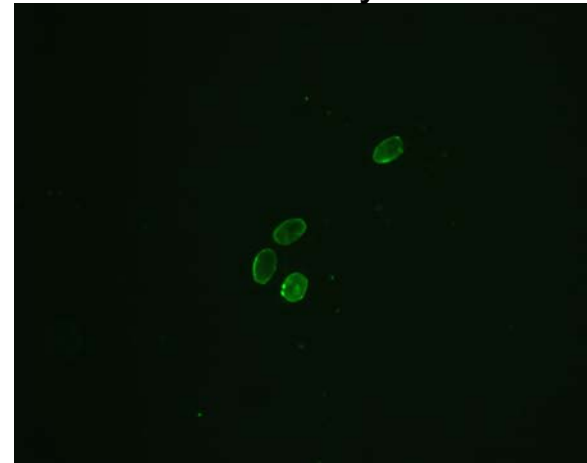
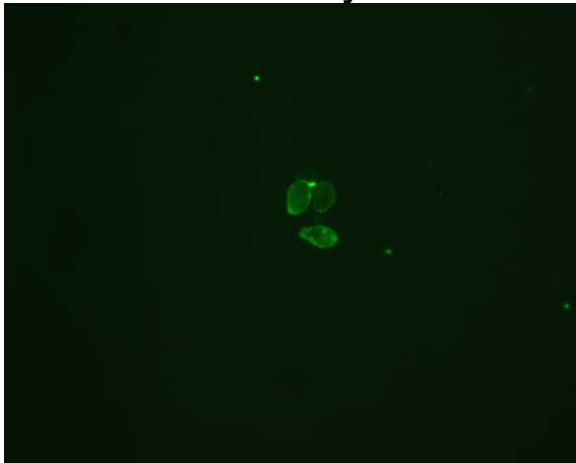


Reaction with *P. pachyrhizi* and *P. meibomiaae*

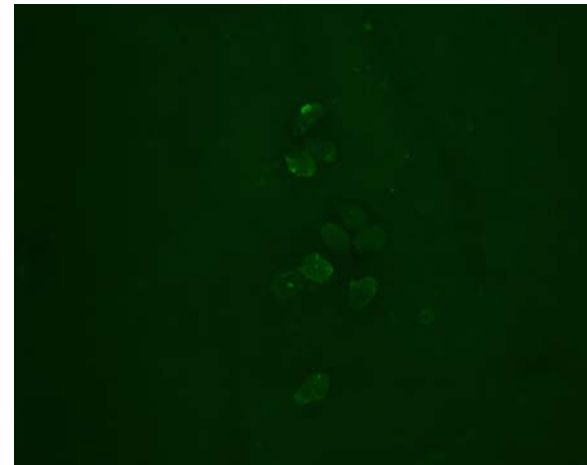
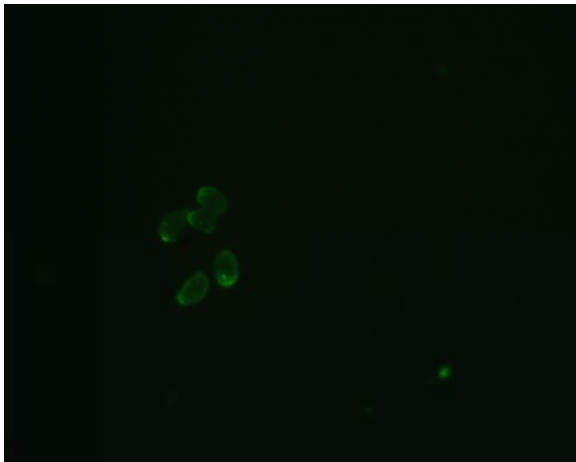
1° Antibody 1A

1° Antibody 2

P. pacchryzi



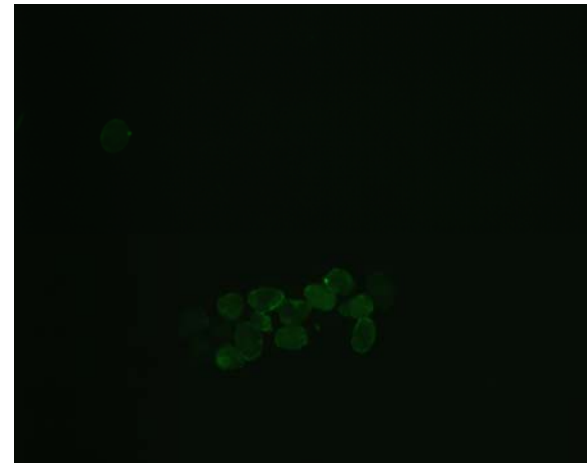
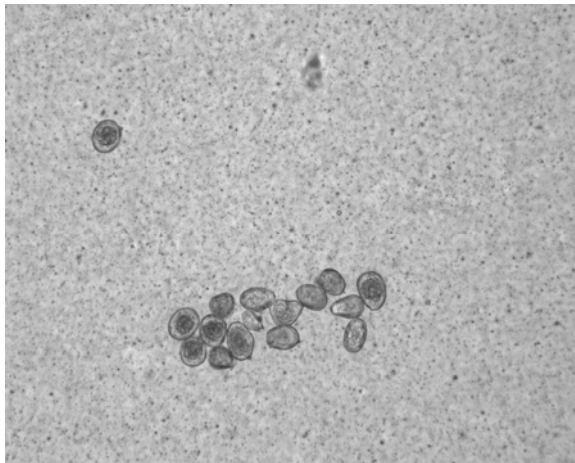
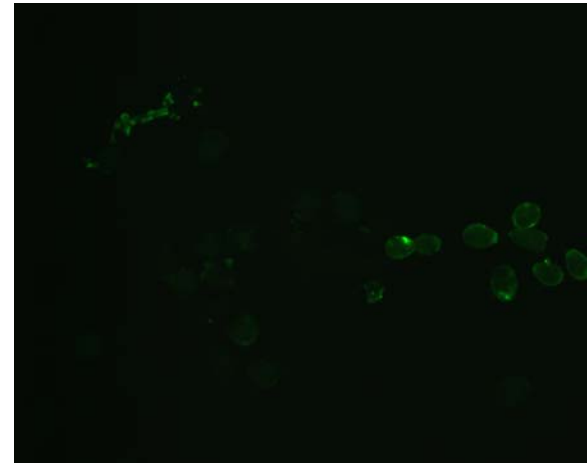
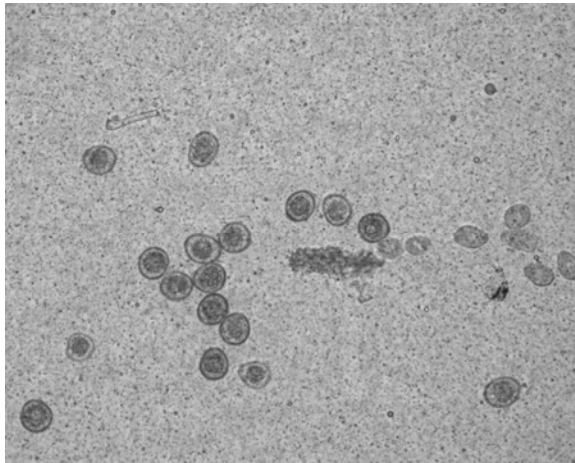
P. meibomiaae



1:500 1:500

Antibody Specificity

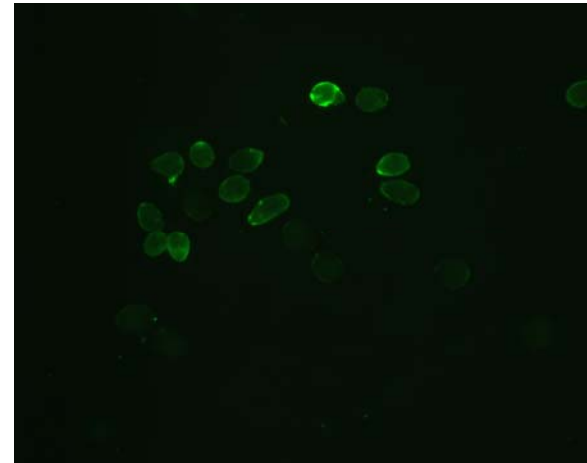
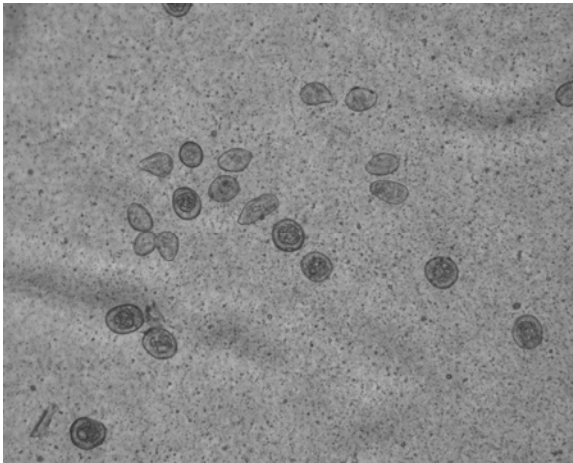
P. pachyrhizi and *P. graminis* with 1° AB 1A (1:500 1:500)



P. meibomiae and *P. graminis* with 1° AB 1 (1:500 1:500)

Antibody Specificity

P. pachyrhizi and *P. graminis* with 1° AB 2 (1:500 1:500)



P. meibomiae and *P. graminis* with 1° AB 2 (1:500 1:500)

Assay Adaptation - Spore Traps



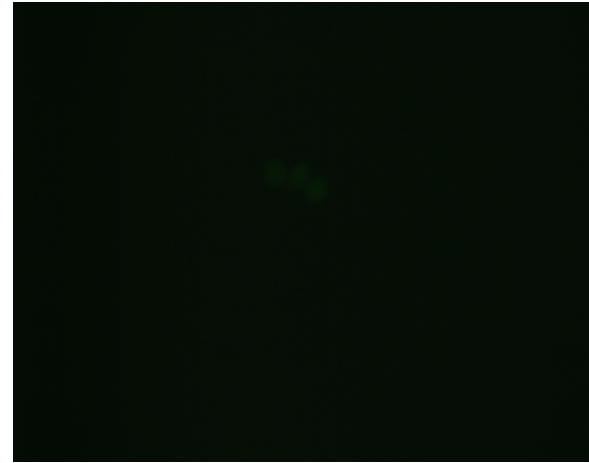
- Original assay parameters worked out using Pp spores in microfuge tubes
- Spore traps capture urediniospores on vaseline-coated slides

Double-stick tape

P. pachyrhizi with 1° AB 1A



P. pachyrhizi without 1° AB

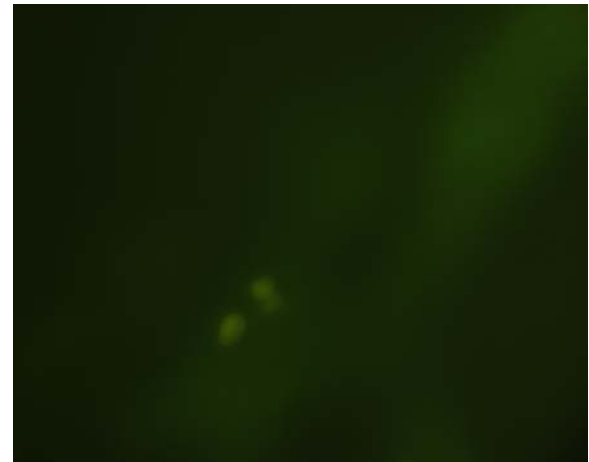


P. pachyrhizi with 1° AB 1A



Vaseline

P. pachyrhizi without 1° AB



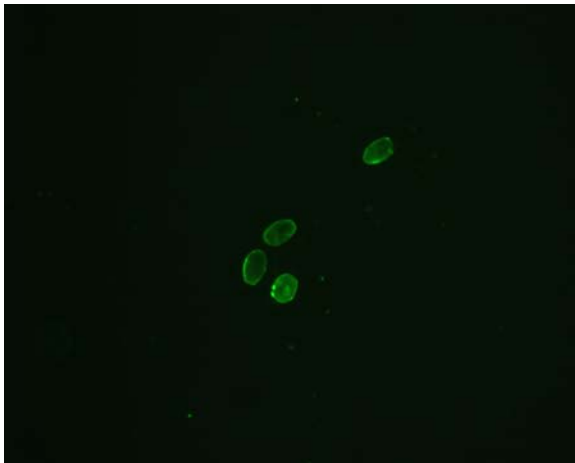


“Stickiness” of *Phakopsora* Spores to Slides

Vaseline			DoubleŠstick tape		
Slide	Initial # spores	% lost spores	Slide	Initial # spores	% lost spores
A	155	14.4	A	144	4.16
B	125	26.7	B	118	3.39
C	131	26.1	C	134	7.46
D	136	26.4	D	128	4.69
E	34	17.7	E	140	3.57
Mean		22.2	Mean		4.7

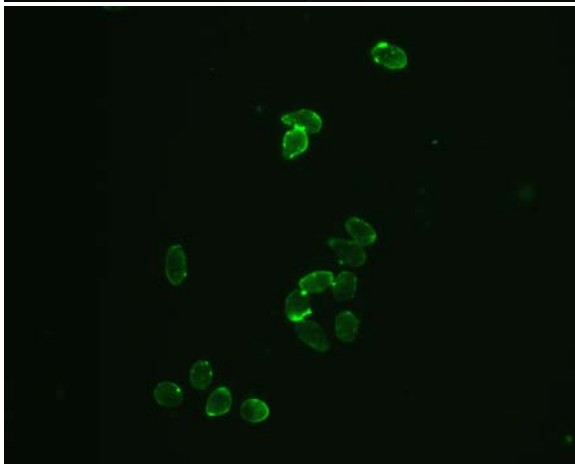
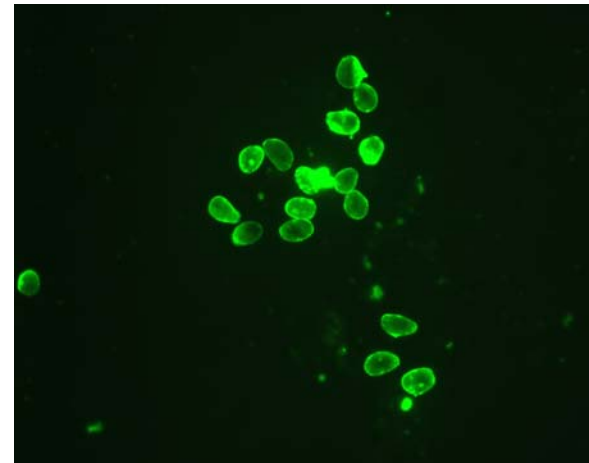
SBR Immunofluorescence Assay Optimization

1:500 1:500

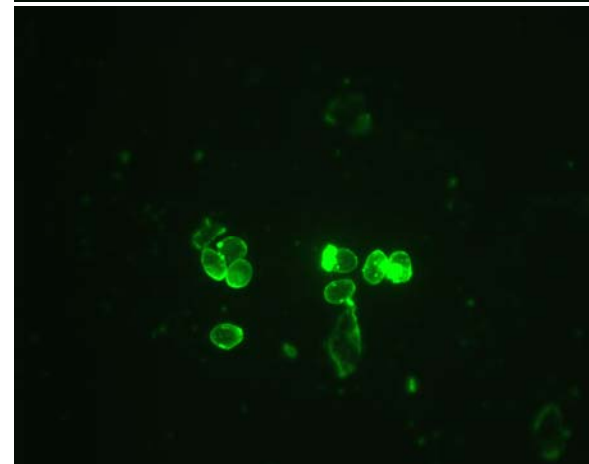


1° AB 2

1:250 1:250

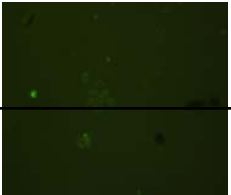
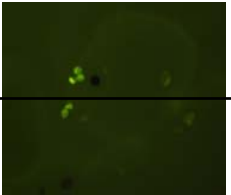


1:500 1:100



1:100 1:100

Field Test Results

Slides name	Date	Slide type	1° Antibody	2° Antibody	SBR- like spores (fluorescent)	
NW (003)	No date	Double sided tape	1A	Alexa Fluor 488	+	
NW (004)	09.05.2006-09.11.2006	Double sided tape	1A	Alexa Fluor 488	+	
NW (6)	09.05.2006	Double sided tape	1A	Alexa Fluor 488	-	
NW 1 (Wood County)	8.21.2006-8.23.2006	Double sided tape	1A	Alexa Fluor 488	-	



Next steps

- Antisera will be available to interested parties
- Complete monoclonal antisera
 - Recombinant secreted proteins (ARS)
 - Polyclonal for the secreted proteins is quite active
- Develop ELISA formats – other formats
- Develop and assess field sampling protocols



Acknowledgments

- This project was funded by soybean check-off dollars from the Iowa Soybean Promotion Board and United Soybean Board
- Antiserum and protocol can be obtained by contacting Anne Dorrance:
dorrance.1@osu.edu